

Thaw and Culture Details

Cell Line Name	WA07					
WiCell Lot Number	WB34437					
Parent Material	WA07-MCB-05					
Provider	University of Wisconsin - Laboratory of Dr. James Thomson					
Banked By	WiCell					
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 1 well of a 6 well plate. WiCell recommends thawing using ROCK Inhibitor for best results.					
Culture Platform	Feeder Dependent					
	Medium: hESC Medium (KOSR)					
	Matrix: MEF					
Protocol	WiCell Feeder Dependent Protocol					
Passage Number	p27 These cells were cultured for 26 passages prior to freeze. WiCell adds +1 to the passage number to best represent the overall passage number of the cells at thaw.					
Date Vialed	17-May-2016					
Vial Label	WA07 p27 WB34437					
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.					

Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation and recoverable attachment after passage	Pass
Identity by STR	UW Translational Research Initiatives in Pathology Laboratory	PowerPlex 16 HS System by Promega	Consistent with known profile	Pass
Sterility	Biotest Laboratories	ST/07	Negative	Pass
Mycoplasma	WiCell	SOP-QU-004	Negative	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Expected karyotype	Pass

Approval Date	Quality Assurance Approval
15-August-2016	A15,0016 X JKG WG Quality Assurance Signed by Gey Janna



Short Tandem Repeat Analysis

WiCell®
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Department of Pathology and Laboratory Medicine TRIP Laboratory (Molecular) http://www.pathology.wisc.edu/research/trip

Sample Report: 11710-STR

Sample Name on Tube: 11710-STR

53.5 ng/μL, (A260/280=1.90)

Sample Type: Cells

Cell Count: ~2 million cells

Requestor: WiCell Research Institute Quality Department Sample Date: N/A Receive Date: 07/11/16 Assav Date: 07/15/16

File Name: STR 160719 wmr

Report Date: 07/21/16

STR Locus	R Locus STR Genotype Repeat #				
FGA	16–18,18.2,19,19.2,20,20.2,21,21.2,22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2,45.2, 46.2	21,22			
TPOX	6-13	8,11			
D8S1179	7-18	13,14			
vWA	10-22	14,15			
Amelogenin	X,Y	X,X			
Penta D	2.2, 3.2, 5, 7-17	13,15			
CSF1PO	6-15	12,12			
D16S539	5, 8-15	12,13			
D7S820	6-14	10,11			
D13S317	7-15	11,12			
D5S818	7-16	11,13			
Penta E	5-24	11,13			
D18S51	8-10, 10.2, 11-13, 13.2, 14-27	12,15			
D21S11	24,24.2,25,25.2,26-28,28.2,29,29.2, 30, 30.2,31, 31.2,32,32.2,33,33.2, 34,34.2,35,35.2,36-38	30,31.2			
TH01	4-9,9.3,10-11,13.3	6,6			
D3S1358	12-20	15,16			

<u>Results:</u> Based on the 11710-STR cells submitted by WiCell QA dated and received on 07/11/16, this sample (Label on Tube: 11710-STR) exactly matches the STR profile of the human stem cell line WA07 comprising 28 allelic polymorphisms across the 15 STR loci analyzed.

<u>Interpretation:</u> No STR polymorphisms other than those corresponding to the human WA07 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 11710-STR sample submitted corresponds to the WA07 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells.

<u>Sensitivity:</u> Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~2-5%.

X RMB Digitally Signed on 07/21/16

X WMR Digitally Signed on 07/21/16

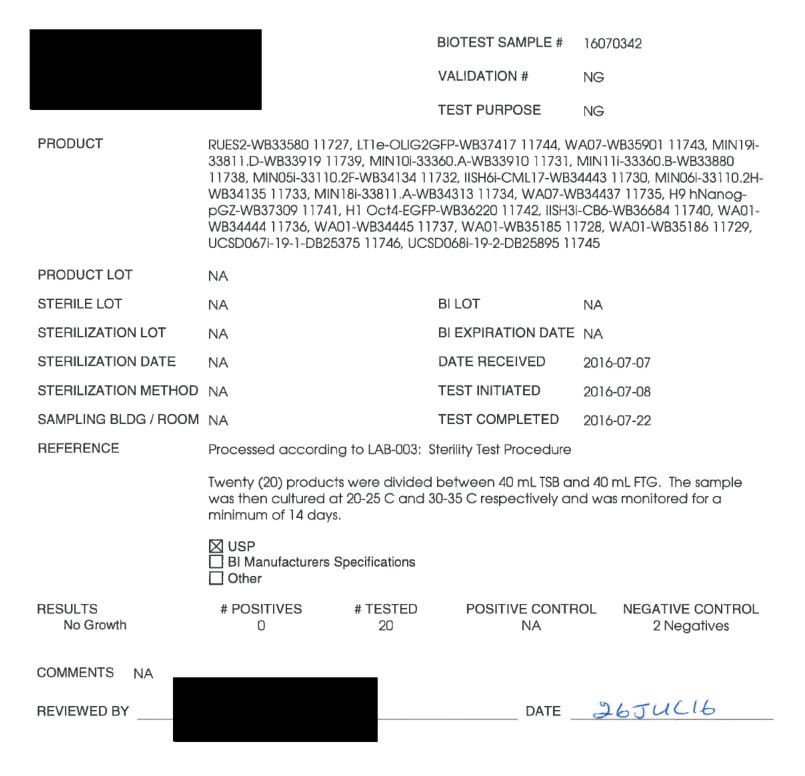
PhD, Director / Co-Director

TRIP Laboratory, Molecular

UWHC Molecular Diagnostics Laboratory / UWSMPH TRIP Laboratory

Sterility Report





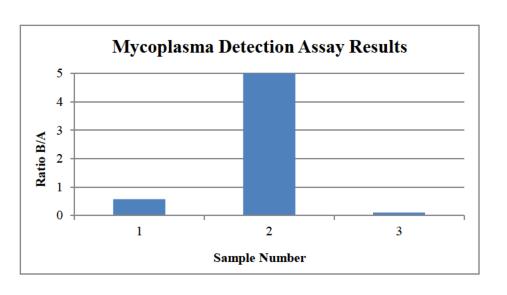
Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. Liability is limited to the costs of the tests,



Mycoplasma Detection Assay Report

Testing Performed by WiCell Lot Release Testing July 8th, 2016 FORM SOP-QU-004.01 Version F Edition 01 Reported by: SM Reviewed by: JB BD Monolight 539

		Read	ing A	A	Read	ing B	В	Ratio		
#	Sample Name	RLU1	RLU2	Ave	RLU1	RLU2	Ave	\mathbf{B}/\mathbf{A}	Result	Comments/Suggestions
1	WA07-WB34437 11710	303	291	297	174	164	169	0.57	Negative	
2	Positive (+) Control	241	239	240	18462	18418	18440	76.83	Positive	
3	Negative (-) Control	340	325	332.5	34	31	32.5	0.10	Negative	





Chromosome Analysis Report: 039745

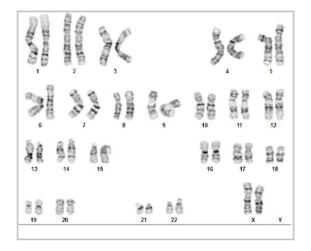
Date Reported: Thursday, June 30, 2016

Cell Line: WA07-WB34437 11710

Passage#: 30

Date of Sample: 6/27/2016

Specimen: hESC Results: 46,XX



Cell Line Gender: Female

Reason for Testing: Lot release testing

Investigator:

Cell: 50 Slide: 2

Slide Type: Karyotype

Total Counted: 20 Total Analyzed: 8

Total Karyogrammed: 4 Band Resolution: 450 - 475

Interpretation:

This is a normal karyotype. No clonal abnormalities were detected at the stated band level of resolution.

Completed by:	CG(ASCP)
Reviewed and Interpreted by:	PhD, FACMG

A signed copy of this report is available upon request.

Date:	Sent By:	Sent To:	QC Review By:
Limitations: This assay allows for microscopic v	visualization of numerical	and structural chromosome abnormalitie	es. The size of structural abnormality that can be detect

is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.

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